NOTE

A SIMILAR ACTION TO UV-IRRADIATION AND A PREFERENTIAL INHIBITION OF DNA SYNTHESIS IN E. COLI BY ANTITUMOR PLATINUM COMPOUNDS

MIKIO SHIMIZU* and BARNETT ROSENBERG

Biophysics Department, Michigan State University, E. Lansing, Michigan 48823, U.S.A.

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Cis-dichlorodiammineplatinum (II) [cis-Pt (II) (NH₃)₂Cl₂], and related compounds were first reported by ROSENBERG *et al.*^{1,2}) to have potent antitumor activities against sarcoma 180 and leukemia 1210 in mice. Subsequently, these compounds were reported effective against DUNNING ascites leukemia and WALKER 256 carcinosarcoma in rats⁸) and in other systems.^{4~6}

Cis-Pt (II) $(NH_3)_2Cl_2$ also causes inhibition of cell division and elongation of bacterial cells,^{7,8)} prophage induction in bacteria,⁹⁾ inhibition of DNA synthesis in mammalian cells,⁴⁾ cross-linking of DNA *in vivo*¹⁰⁾ and *in vitro*,¹¹⁾ and immunosuppression in mice.^{12,13)} These activities of *cis*-Pt (II) $(NH_3)_2Cl_2$ are very similar to those of difunctional alkylating agents. Bleomycin was reported to differ in mode of action from radiation even though it caused bacterial elongation and inhibition of DNA synthesis.^{14,15)} This study was performed to determine the susceptibilities of four isogenic sub-strains of

Escherichia coli-Bs-1, Bs-2, B and B/r¹⁶⁾ to UV irradiation, *cis* and *tran*-Pt (II) $(NH_8)_2Cl_2$, mitomycin C, bleomycin, rifamycin and chloramphenicol. *In vitro* minimum inhibitory concentrations were determined by the dilution method. Overnight bacterial cultures in nutrient broth (Difco) were added by small capillaries to nutrient broth agar plates which contained two-fold dilutions of drug. After incubation for 16 hours at 35°C, plates were examined to determine the minimum concentration of drug which prevented visible growth of bacteria.

The effects of UV and drugs on the viability of E. coli strains were determined by the following procedures: Each strain, grown logarithmically in a medium $((NH_4)_2HPO_4 2.5 g, KH_2PO_4 1.5 g,$ $MgSO_4 \cdot 7H_2O$ 0.1 g, sodium glutamate 3.0 g, glucose 3.0 g per liter), was harvested, washed and resuspended in a saline or in this medium without glucose. To determine UV-sensitivity of strains, each cell suspension $(5 \times 10^8 \text{ cells/ml})$ in saline, 10 ml/petri dish of 10 cm diameter) was exposed for 30 seconds to a 10-w UV germicidal lamp at a distance of 30 cm. Aliquots of the suspensions were plated out and incubated. Surviving cells were expressed relative to numbers in controls not irradiated. To determine lethality of drugs to these strains, various concentrations of drugs were added to each cell suspension (10⁹ cells/ml in the medium without glucose). The mixtures were incubated at 37°C for 30 minutes. Dilutions of these sus-

	Minimum inhibitory concentration (µм)			
	E. coli Bs-1	E. coli Bs-2	E. coli B	<i>E. coli</i> B/r
UV*	3×10 ⁻¹⁰	3×10-7	0.2	1.0
Cis-Pt (II) (NH ₃) ₂ Cl ₂	38	38	150	300
Trans-Pt (II) (NH ₃) ₂ Cl ₂	> 300	> 300	> 300	> 300
Mitomycin C	0.1	0.02	0.4	3.3
Bleomycin	2,2	0.6	1.1	9.0
Rifamycin	6.3	3.2	6.3	12.5
Chloramphenicol	2.0	2.0	2.0	2.0

Table 1. The sensitivity of the E. coli strains to drug.

* Cells were exposed for 30 seconds to a 10-w UV lamp at a distance of 30 cm and figures indicate the survivors relative to unirradiated controls.

* Present address: Tokyo Research Laboratory, Kyowa Hakko Kogyo Co., Ltd., Tokyo, Japan.

Fig. 1. Bacteriocidal effects of *cis*-Pt (II) (NH₃)₂Cl₂ and mitomycin C on UV-sensitive strains and UV-resistant strains.

Each strain, grown logarithmically in a medium $((NH_{4})_2HPO_4 2.5g, KH_2PO_4 1.5g, NaCl 5.0g, MgSO_4.7H_2O 0.1g, sodium gulta$ mate 3.0g, glucose 3.0g per liter), was harvested, washed and resuspended in the medium without glucose to a concentration of10⁹ cells/ml. Each concentration of drug was added to each cellsuspension. The mixture was incubated at 37°C for 30 minuteswithout shaking. After appropriate dilutions, the suspension wasspread on nutrient agar plates. The colonies were counted afteran overnight incubation of the plates.



Fig. 2. Effects of *cis*-Pt (II) (NH₃)₂Cl₂ and mitomycin C on nucleic acids and protein synthesis by *E. coli* Bs-1.

The experiment was performed by the method described by SEKIGUCHI *et al.*¹⁷⁾ Briefly, bacterial cultures containing various levels of drug were incubated with shaking at 37°C for 30 minutes. After collecting cells by centrifuge, nucleic acids were extracted by heating cells in 0.5 N perchloric acid at 90°C for 15 minutes. DNA and RNA in the extracts were determined by diphenylamine reaction and by orcinol reaction, respectively. The residue after extraction was dissolved in 1 N NaOH, and protein was determined by the method of LOWRY *et al.*



pensions were spread on nutrient broth agar plates, incubated overnight and surviving cells were tabulated.

Table 1 shows that UV-sensitive mutants, E. coli Bs-1 and E. coli Bs-2, were much more sensitive to UV irradiation than the parent E. coli B strain. Table 1 and Fig. 1 show that cisPt (II) $(NH_3)_2Cl_2$ and mitomycin C were more effective on E. coli Bs-1 and E. coli Bs-2 than on E. coli B and the UV-resistant mutant E. coli B/r. Bleomycin, as reported by ENDO,¹⁴⁾ showed no significant difference in its antibacterial activity on UVsensitive strains and the parent strain, but it was less active against the UV-resistant mutant *E. coli* B/r (Table 1). Rifamycin and chloramphenicol exhibited no significant differences in minimal inhibitory concentrations against UV-sensitive and UV-resistant strains (Table 1).

The effects of cis-Pt (II) $(NH_3)_2Cl_2$ and mitomycin C on nucleic acids and protein synthesis by E. coli Bs-1, were determined essentially by the method of SEKIGUCHI et al.17) Bacterial cultures containing various concentrations of drug were incubated with shaking at 37°C for 30 minutes. Cells were separated by centrifugation and nucleic acids in the cells were extracted by heating in 0.5 M perchloric acid at 90°C for 15 minutes. DNA and RNA in the extracts were determined by diphenylamine and by orcinol reactions, respectively. The residues after perchloric acid extraction were dissolved in 1 N NaOH, and proteins were determined by the method of LOWRY et al. Fig. 2 shows that cis-Pt (II) $(NH_3)_2Cl_2$ and mitomycin C caused a preferential inhibition of DNA synthesis in bacterial cells as was observed with mammalian cells.4)

These results suggest that cis-Pt (II) (NH₃)₂Cl₂ is very similar in mode of action to difunctional alkylating agents.

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